ENANTIOSPECIFIC SYNTHESIS OF C₂₀-C₂₈ SEGMENT OF CONCANAMYCIN A: APPLICATION OF DIETHYLISOPROPYLSILYL PROTECTING GROUP

Kazunobu Toshima, Mari Misawa, Kazumi Ohta, Kuniaki Tatsuta,* and Mitsuhiro Kinoshita* Department of Applied Chemistry, Keio University, 3-14-1, Hiyoshi, Kohoku-Ku, Yokohama 223, JAPAN

<u>Summary</u>: A complex C_{20} - C_{28} segment 5 of concanamycin A (1) has been first synthesized without side reactions by effective de-O-silylation of 2a, which was constructed from the ethyl ketone 3 by glycosidation with the fluoride 4.

The antibiotic concanamycin A (1) is a major component of concanamycins which were isolated in 1984 by Kinashi et al¹) and belongs to a new class of 18-membered macrolide with a long side chain which incorporates a six membered hemiketal ring having 2-deoxy- β -D-rhamnose moiety¹). Concanamycin (1) was identified with the antifungal antibiotics, folimycin and A666-I and inhibits the proliferation of mouse splenic lymphocytes stimulated by concanavalin A¹). The absolute configuration of 1 has been established by X-ray crystallographic analysis of its diacetate derivative²). In this article we would like to report the first enantiospecific synthesis of 5 which is a promising C₂₀-C₂₈ segment of concanamycin A, and usefulness of diethylisopropylsilyl (DEIPS) group^{3,4}) as an effective O-protecting group, especially in deprotection of 2a.

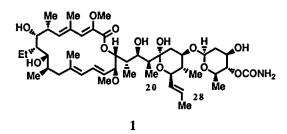
The construction of 2a would be achieved by the glycosidation of the ethyl ketone segment 3 with the sugar segment 4, both of which could be enantiospecifically synthesized from appropriate carbohydrate derivatives. The selective formation of β -glycosidic linkage of 2-deoxy-D-sugar would become formidable task⁵) and the selection of the protecting groups, R¹ and R² should naturally become the key problem in the final stage of the conquest of 5 and 1. Considering these points, we started the construction of 3. The starting material 6⁶) was treated with acetone and catalytic amount of BF₃•Et₂O (0.6 equiv)³) at 26°C for 17h to afford 7⁷) ([α]_D +21° (c 1.26, CHCl₃)) in one step in 63% yield. Selective 5,6-de-O-isopropylidenation of 7 with 75% AcOH at 30°C for 2.5h gave diol 8⁷) ([α]_D +15° (c 0.62, CHCl₃)) in 72% yield. Periodate-oxidation (NaIO₄, acetone-H₂O, 26°C, 1h) of 8, followed by the Wittig reaction of the generated aldehyde with 1.5 equiv. of methoxycarbonylmethylenetriphenylphosphorane at 25°C for 2h afforded desired trans product 9⁷) ([α]_D +52° (c 1.30, CHCl₃)) with high selectivity in 77% overall yield. The Wittig product 9 was treated with DIBAL in toluene under ice-cooling for 0.5h to give

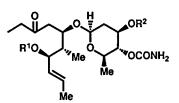
the crude allylalcohol **10** which was converted (MsCl, LiBr, Et₃N, CH₂Cl₂, 25°C, 8h) into bromide **11**⁷) ([α]₅₄₆ +0.8° (c 1.20, CHCl₃)) in 81% overall yield from **9**. LAH-Reduction (ether, 26°C, 4h) of **11** proceeded cleanly to afford **12**⁷) ([α]_D +11° (c 1.50, CHCl₃)) in 92% yield. Hydrolysis of **12** (50% AcOH-H₂O, 80°C, 0.5h) followed by LAH-reduction (THF, 70°C, 2.5h) afforded the triol, which was subjected to the selective protection of the 1,2-diol with carbonate (1.3equiv *N*,*N*′-carbonyldiimidazole, benzene, 26°C, 2h) and the resulted mono-alcohol **13**⁷) (mp 62.5-63.5°C, [α]_D -45° (c 1.38, CHCl₃)) was silylated (DEIPSCl, imidazole, CH₂Cl₂, 25°C, 2h) to give **14**⁷) (IR(CHCl₃) v_{max} 1800_{cm}⁻¹, [α]_D -46° (c 0.98, CHCl₃)) in 75% overall yield from **12**. Hydrolysis (NaOH, MeOH, 0°C, 2.5h) of **14** followed by selective mono-tosylation (2.5 equiv TsCl, Py, 25°C, 4h) of the primary alcohol and epoxidation (NaOMe-MeOH, CHCl₃, 25°C, 1.5h) gave **15**⁷) ([α]_D -10° (c 1.14, CHCl₃)) in 84% yield from **14**. Reaction of **15** with 2-ethyl-2-lithio-1,3-dithiane (5 equiv) (THF, -20°C, 1h) afforded the crude **16** whose dithioacetal group was cleaved (1:1 HgCl₂-HgO, 80% acetone-H₂O, 0°C, 0.5h) to give the pure ketone **3**⁷) (IR(CHCl₃) v_{max} 1709 cm⁻¹, [α]_D +6.9°, (c 0.46, CHCl₃)) in 72% overall yield from **15**.

2-Deoxy-D-glucose was selected as the starting material for the construction of **4**. Benzyl glycosidation (BzlOH, BF₃•Et₂O, 80°C, 3h) of 2-deoxy-D-glucose, followed by selective iodonation (1.4 equiv I₂, 1.5 equiv PPh₃, 3.0 equiv imidazole, 70°C, 2h) afforded **17**⁷) which was subjected to Bu₃SnH-reduction (AIBN, 80°C, 3h) to give **18**⁷) in 71% overall yield from 2-deoxy-D-glucose. Mono silylation (1.2 equiv DEIPSCI, 1.3 equiv imidazole, CH₂Cl₂, 25°C, 3h) proceeded cleanly with high selectivity to afford **19**⁷) whose free alcohol was carbamoylated (1. Chloroacetyl isocyanate, CH₂Cl₂, 0°C, 1h; 2. Zn, MeOH, 25°C, 5h)⁸) to afford **20**⁷) in 88% overall yield from **18**. **20** was hydrogenerated by H₂ in the presence of Pd(OH)₂ in dioxane at 25°C for 4h to give **21**⁷) in 97% yield. Conversion of the free sugar **21** into the fluoride was accomplished by using 1.2 equiv. of DAST⁹) in THF-ether (2:1) (-30 → +25°C, 20 min) to afford **4**⁷) (mp 103.5-104.5°C, [α]_D +21°, (c 1.10, CHCl₃)) in 86% yield (α/β =14/1).

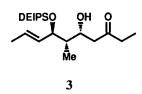
The glycosidation of **4** to the hydroxy group of **3** was best effected by carefully modified Mukaiyama method¹⁰⁾ (1 equiv **3**, 1 equiv **4**, 1 equiv SnCl₂, MS 4A, CH₂Cl₂, -50°C, 3h then -40°C, 5h) to give the desired β -glycoside **2a**⁷⁾ (mp 141-142°C, [α]_D -15°, (c 1.00, CHCl₃)) with moderate selectivity in 30 % yield. By the procedure described in the synthesis of **2a**, the another segment **2b**⁷⁾ (mp 150-151°C, [α]_D -11°, (C 0.82, CHCl₃)) was also prepared by using only TBDMS-Cl as the silylating agent. In the deprotection of **2a** and **2b**, mild acidic condition was used without side reactions instead of tetrabutylammonium fluoride as the deprotection reagent because of, their labile β -ketol type structure³,11). Treatment of **2b** with AcOH:1%KF•HF(aq):THF (3:1:3) did not give the desired **5** even at 30°C for 46 h but gave **22**⁷⁾ in 94% yield. In contrast to this fact, the desilylation of **2a** by AcOH:1%KF•HF(aq):THF (3:1:3) (30°C, 46h) proceeded smoothly to afford **5**⁷) (mp 170-171°C, [a]_D -29°, (c 0.72, MeOH)) in 83% yield.

Thus C_{20} - C_{28} segment 5 of concanamycin A which is a promising key intermediate for the synthesis of 1 has been first synthesized by effective use of DEIPS group which was recently developed³⁾ and characterized⁴⁾ in our laboratories. Improvement of the glycosidation of 4 with 3 is now under study.



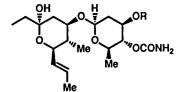


2a: $R^1 = R^2 = DEIPS$ **2b:** $R^1 = DEIPS$, $R^2 = TBDMS$

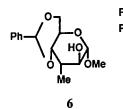


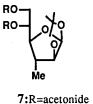


4:R=DEIPS

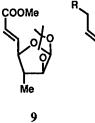


5:R=H 22:R=TBDMS





7:R=acetonide 8:R=H

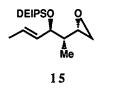


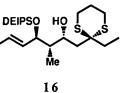
RO Q H Me

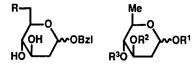
10:R=OH 11:R=Br 12:R=H

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13:R=H 14:R=DEIPS







17:R=I **19:**R¹=Bzl, R²=DEIPS, R³=H **18:**R=H **20:**R¹=Bzl, R²=DEIPS, R³=CONH₂ **21:**R¹=H, R²=DEIPS, R³=CONH₂ <u>Acknowledgement:</u> We are grateful to the Institute of Microbial Chemistry for the generous support of our program. Financial support by the Ministry of Education, Science and Culture (Grant-in-Aid Scientific Research) is deeply acknowledged. We also thank Messrs. S. Nishimura and A. Asai for their technical assistance.

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- 7). All compounds were purified by silica gel column chromatography and were fully characterized by spectroscopic means and elemental analyses. Optical rotations were mesured using a 0.2dm or 0.5dm tube at 25°C. Melting points were uncorrected. Significant ¹H-NMR spectra [δ (TMS), J(Hz)] are the following. 3: (270MHz, CDCl₃) 0.55-0.7 (4H, m), 0.94 (3H, d, J=7.8), 0.95-1.05 (13H, m), 1.06 (3H, t, J=7.6), 1.51 (1H, ddt, J=7.8, 4.4, 2.0), 1.71 (3H, d, J=4.8), 2.32 (1H, dd, J=15.6, 4.0), 2.49 (2H, dq, J=7.6, 2.0), 2.69 (1H, dd, J=15.6, 7.2), 3.52 (1H, br s, OH), 4.12(1H, ddd, J=4.4, 4.4, 1.2), 4.51 (1H, ddd, J=7.2, 4.0, 2.0), 5.45-5.7 (2H, m). 4: (270MHz, CDCl₃) 0.55-0.7 (4H, m), 0.9-1.05 (13H, m), 1.25 (3H, d, J=6.4), 1.81 (1H, dddd, J=40.4, 13.8, 11.0, 2.4), 2.28 (1H, dddd, J=13.8, 9.8, 6.0, 1.6), 3.91 (1H, dq, J=10.0, 6.4), 4.07 (1H, ddd, J=11.0, 10.0, 6.0), 4.54 (1H, dd, J=10.0, 10.0), 4.65- 4.8 (2H, m), 5.63 (1H, ddd, J=51.8, 2.4, 1.6). 2a: (270MHz, CDCl₃) 0.5-0.7 (8H, m), 0.78 (3H, d, J=6.8), 0.9-1.05 (14H, m,), 1.04 (3H, t, J=7.2), 1.15-1.3 (1H, m), 1.21 (3H, d, J=6.2), 1.5-1.65 (1H, m), 1.68 (3H, dd, J=6.2, 1.6), 1.95 (1H, ddd, J=12.8, 5.4, 1.8), 2.35-2.55 (3H, m), 2.77 (1H, dd, J=15.8, 8.4), 3.26 (1H, dq, J=9.6, 6.2), 3.71 (1H, ddd, J=9.8, 8.6, 5.4), 4.13 (1H, dd, J=8.2, 8.2), 4.39 (1H, dd, J=9.8, 9.8), 4.45-4.55 (1H, m), 4.52 (1H, dd, J=10.0, 10.0), 4.64 (2H, br s), 5.35 (1H, ddq, J=15.6, 8.2, 1.6), 5.55 (1H, dq, J=15.6, 6.2). 5: (250MHz, pyridine-d5) 1.13 (3H, t, J=7.8), 1.17 (3H, d, J=6.3), 1.42 (3H, d, J=6.0), 1.5-1.7 (2H, m), 1.62 (3H, d, J=4.8), 1.94 (2H, q, J=7.8), 2.05-2.25 (1H, m), 2.45-2.6 (2H, m), 3.50 (1H, dq, J=10.0, 6.0), 4.05-4.2 (1H, m), 4.25-4.4 (2H, m), 4.7-4.85 (2H, m), 4.89 (1H, dd, J=9.8, 1.3), 5.01 (1H, dd, J=9.5, 9.5), 5.5-5.75 (2H, m).
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